INTRODUCTION

American tegumentary leishmaniasis (ATL) is a group of zoonotic diseases widespread in 22 countries from Texas (USA) to northern Argentina. It is caused by flagellar protozoa belonging to the genus Leishmania in which a tremendous diversity or polymorphism has been found. The diseases caused by different species are distinct in their clinical manifestations, drug responses, and prognosis. Therefore, it is necessary to characterize and identify specifically the parasites obtained in each endemic area where a survey is taken.

For this purpose, multilocus enzyme electrophoresis (MLEE) analysis has been recommended by the World Health Organization (WHO).

In northern Argentina, some areas have been reported to be endemic for human and canine leishmaniasis. However, little is known about the causative agents, Leishmania spp., and the transmission cycles implicated there. Salta and Corrientes Provinces are endemic areas, where most of the clinical forms of human ATL showed single and multiple cutaneous (CL), mucocutaneous (MCL), or mucosal lesions (Borda CE and others, unpublished data). Vegetative nonulcerated lesions with metastasis to the lymph nodes and viscus (spleen) have also been described. As to the causative parasites, it was found that L. (Viannia) braziliensis was predominant in Salta Province, followed by L. (Leishmania) amazonensis, and L. (V.) guyanensis, though relatively few isolates were specifically characterized to date.

Previously, we studied the clinical, parasitological, and serological characteristics of canine tegumentary leishmaniasis in Salta Province. Several observations from these works suggested that dogs might be considered as a susceptible host without having a principal role in the spread of the disease. However, any Leishmania stock was not isolated from dogs in this area before.

This paper reports the species attribution and the examinations of the relationship between Leishmania isolates from different clinical forms of human and canine ATL in northern Argentina, based on MLEE data; the first description of canine Leishmania isolates in the country is herein provided. The relevance of the intraspecific polymorphism to the induction of different clinical forms, as well as its incidence in the host-reservoir concept, is briefly discussed.

MATERIALS AND METHODS

Parasites and study area. Sixteen stocks of Leishmania were included in this study. Fourteen of them, from 11 humans, and one stock from a dog were isolated in Salta Province, northwest Argentina. The remaining one was obtained from a patient in Corrientes Province, located in the northeast of Argentina. Their denomination, the clinical data of the host, and a description of the geographical origin are shown in Table 1 and Figure 1. The samples were taken from patients who were accessed in the four institutions, viz., the Instituto de Investigaciones en Enfermedades Tropicales, Subsede Orán, Universidad Nacional de Salta; the Dermatology Ward of Hospital del Milagro; the Dermatology and Otorhinolaryngology Wards of Hospital San Bernardo of Salta City; and Centro Nacional de Parasitología y Enfermedades Tropicales, Universidad Nacional del Nordeste, Corrientes City. A part of the current isolates was obtained from humans and canine cases during house-to-house visits.

All the referred patients, from whom Leishmania parasites were isolated from their lesions, were systemically treated for 25–30 days with 10–20 mg d⁻¹ kg body wt⁻¹ of pentavalent antimony for each cycle. In the cases of the incomplete clinical cure of lesion, another cycle of treatment or amphotericin B treatment was given. The clinical controls and treatment protocols were exclusively conducted for local physicians. The patients voluntarily consented to participate in this study, and the Bioethical Commission of Health Ministry of Salta Province approved the procedures.

Six WHO reference strains were included; L. (V.) braziliensis, MHOM/BR/75/M2904; L. (V.) panamensis, MHOM/PA/71/LS94; L. (V.) guyanensis, MHOM/BR/75/M4147; L. (L.) amazonensis, MHOM/BR/73/M2269; L. (L.) mexicana,
<table>
<thead>
<tr>
<th>Zymodemes and species assignment of 16 Argentinian Leishmania stocks and clinical-epidemiological aspects of the hosts</th>
<th>Age (years)</th>
<th>Geographical origin</th>
</tr>
</thead>
<tbody>
<tr>
<td>KMS1</td>
<td>20 M 30</td>
<td>Salta, Chiquim, Sola</td>
</tr>
<tr>
<td>KMS2</td>
<td>30 M 40</td>
<td>Salta, Chiquim, Sola</td>
</tr>
<tr>
<td>KMS3</td>
<td>40 M 50</td>
<td>Salta, Chiquim, Sola</td>
</tr>
<tr>
<td>KMS4</td>
<td>50 M 60</td>
<td>Salta, Chiquim, Sola</td>
</tr>
</tbody>
</table>

**Clinical form, lesion features, and evolution**

<table>
<thead>
<tr>
<th>Zymodemes</th>
<th>Species</th>
<th>Lesion</th>
<th>Evolution (days)</th>
<th>Sex</th>
<th>Sexual maturation</th>
<th>Species designation</th>
</tr>
</thead>
<tbody>
<tr>
<td>KMS1</td>
<td>L. (V.) braziliensis</td>
<td>Forehead</td>
<td>Ulcerative-infiltrative plaque</td>
<td>F</td>
<td></td>
<td></td>
</tr>
<tr>
<td>KMS2</td>
<td>L. (V.) braziliensis</td>
<td>Forehead</td>
<td>Ulcerative-infiltrative plaque</td>
<td>M</td>
<td></td>
<td></td>
</tr>
<tr>
<td>KMS3</td>
<td>L. (V.) braziliensis</td>
<td>Forehead</td>
<td>Ulcerative-infiltrative plaque</td>
<td>M</td>
<td></td>
<td></td>
</tr>
<tr>
<td>KMS4</td>
<td>L. (V.) braziliensis</td>
<td>Forehead</td>
<td>Ulcerative-infiltrative plaque</td>
<td>M</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

**Isolation and mass cultivation.** Thirteen stocks were obtained by aspiration of the CL or MCL lesion edges of humans or dogs, using a syringe with 0.5 mL of sterile proline balanced salts solution (PBSS) containing 100 U/mL penicillin and 50 µg/mL streptomycin (P E). The material aspirated was inoculated into Difco blood agar (USMARU) medium containing 20% of defibrinated rabbit blood. One milliliter of PBSS with P E was added after 4 days of incubation at 23°C. A final volume of liquid phase of 2 mL was reached after 4 days more. One stock (JDM1) was obtained by an inoculation of the aspirated material into a footpad of a golden hamster (Mesocricetus auratus). Two months after the inoculation, the footpad tissue was aseptically removed, homogenized, and inoculated into five culture tubes containing 2 mL of sterile liver infusion tryptose medium supplemented with P E and 20% heat-inactivated fetal bovine serum (hiFBS). The remaining two stocks (AZ3, LBC1) were isolated by skin biopsy of the cutaneous lesion edges and transferred to Novy-Nicolle-MacNeal (NNN) medium.

The cultures were examined regularly from the 4th to 30th day of inoculation. When promastigotes were not found, the culture was gently centrifuged and the used USMARU medium was replaced by a fresh one after washing the pellet twice in 5 mL of PBS.

For mass cultivation, 1 mL of promastigotes-culture in logarithmic phase was transferred to 6 mL RPMI 1640 medium supplemented with 10% of hiFBS. The culture volume was duplicated once every 2 days to reach a volume of 70 mL and incubated for a maximum of 12 days. The promastigotes were harvested by centrifugation at 10,000 rpm for 15 minutes at 4°C, after washing (2,500 rpm, 20 minutes) 3 times with sterile 0.85% NaCl solution. The pellets were stored in a deep freezer (−80°C) until use.

**Sample preparation.** The soluble enzyme extracts were obtained by lysis of the pellets in a double volume of stabilizer solution containing 2 mM dithiotreitol, 2 mM e-aminocaproic acid, and 2 mM EDTA. After 3 cycles of freezing and thawing, the supernatants obtained by centrifugation at 27,000 rpm at 4°C were aliquoted and stored at −80°C.12

**Enzyme electrophoresis.** Electrophoresis and staining procedures were performed on cellulose acetate support (Sebia; Moulineux, France) following the methods described by Kreutzer and Christensen and by Evans, with slight modifications.12,14 Pictures of gels were taken with a Kodak EDAS 290 digital camera. Each enzyme extract was analyzed at least twice by 11 enzyme systems for 12 putative enzymatic loci: alanine aminotransferase (E.C.2.6.1.2, ALAT), aspartate aminotransferase (E.C.2.6.1.1, ASAT), glucose-6-phosphate dehydrogenase (E.C.1.1.1.49, G6PDH), 6-phosphogluconate dehydrogenase (E.C.1.1.1.44, 6PGDH), glucose-phosphate isomerase (E.C.5.3.1.9, GPI), malate dehydrogenase (E.C.1.1.1.37, MDH), malic enzyme (E.C.1.1.1.40, ME), mannose-phosphate isomerase (E.C.5.3.1.8, MPI), nucleoside hydrolase (inosine) (E.C.2.4.2.1, NH₄), phosphoglucomutase (E.C.2.7.5.1, PGM), and pyruvate kinase (E.C.2.7.1.40, PK).

**Data analysis.** The distance that each reproducible enzyme band migrated from the origin (anode) was measured. The bands were numerated from the fastest (the nearest one to the cathode) to the slowest for each enzyme gel or zymogram. The obtained set of bands defined an electrophoretic profile for each extract. Stocks with the same profile were grouped in the same zymodeme; the species attribution was made comparing the profiles with the WHO reference *Leishmania* strains. To examine the relationship between the zymodemes, Jaccard distances (JD) were obtained for all pairwise comparisons.15 A distance matrix was built and transformed into a phenogram by the unweighted pair group method using the MEGA version 2.1 Software.16

**RESULTS**

Fourteen stocks, included the isolate from the dog were assigned by MLEE analysis of 12 enzymatic loci to *L. (V.) braziliensis* and two to *L. (V.) guyanensis* (Table 1; Figure 2A). No *L. (L.) amazonensis* was isolated from any patient. For the *L. (V.) braziliensis* stocks, the isolates from humans were grouped in 2 zymodemes, KMS 1 and KMS 2, differing in 1 locus, MDH (Figure 2B). The stock from the dog was assigned to another zymodeme, KMS 3 that differed from KMS 1 only in 1 locus, ME (Figure 2C), and from KMS 2 in 2 loci, ME and MDH (Figures 2B and 2C).

Both *L. (V.) guyanensis* isolates were grouped in the same zymodeme, KMS 4, that included the reference strain. Two loci, G6PDH and 6PGDH, distanced them from the *L. (V.) panamensis* reference strain. The enzymatic profiles are shown in Table 2. The relationship between the zymodemes is shown in a phenogram (Figure 3). Over a JD of 0.9, 2 clusters, corresponding to the subgenera *Vianna* and *Leishmania*, can be distinguished. The zymodemes KMS 1 and KMS 2 were more closely related with each other (JD = 0.056) than with KMS 3 (JD = 0.126) and with *L. (V.) braziliensis* reference strains (JD = 0.161). The JD observed between KMS 4 and *L. (V.) panamensis* reference strains was 0.222.

In three different cases of the current study, two stocks each were isolated from the same patient (MHOM/AR/02/MASS5, MHOM/AR/03/MRO2, and MHOM/AR/03/HNO3); they expressed the same enzymatic profile for each pair, without demonstrating the possibility of co-infection with multiple zymodemes and/or species of *Leishmania* parasite (Table 1; Figure 3).

**DISCUSSION**

*Leishmania* species identification is essential for clinical or epidemiologic surveys because of the diversity found among...
In this work, *L. (V.) braziliensis* was incriminated as a causative agent of three different clinical forms of ATL, viz., CL, MCL, and RCL (recurrent cutaneous leishmaniasis) and involved in failure of treatment with pentavalent antimony drug. *Leishmania (V.) braziliensis* was the predominant species in Salta Province, concordant with previous studies in which the samples were taken mainly from CL cases.

Although many factors are responsible for the clinical expression of *Leishmania* infection, it has recently been reported that intraspecific polymorphism in *L. (V.) braziliensis* species may be related to specific clinical forms of ATL. In the current study, both of the *L. (V.) braziliensis* zymodemes, KMS 1 and KMS 2, were found to have an ability to metastasize to mucocutaneous tissues and lymph nodes. However, we could not exclude the possibility that they have a tropism for different tissues, such as mucosa or skin and lymph nodes. Again, the main zymodeme, KMS 1, was isolated from both MCL and CL cases; the other zymodeme, KMS 2, is composed of isolates that are able to infect the lymph node and induce recurrent cutaneous lesions far from the original scar.

*Leishmania (V.) guyanensis* (zymodeme KMS 4) was isolated from patients with CL in two different geographic regions, Salta in the northwest and Corrientes in the northeast of Argentina. The same zymodeme grouped these stocks and WHO reference strain, confirming the homogeneity previously reported for this species in other areas of different countries.

The current result confirms the presence of this species in Argentina, which was reported for the first time in 1987 from a patient in the same endemic area, Orán, Salta.

Recently, it has been proposed that *L. (V.) guyanensis* and

---

**Table 2**

Enzymatic profiles for Argentinian stocks and WHO reference strain zymodemes

<table>
<thead>
<tr>
<th>Zymodeme</th>
<th>ALAT</th>
<th>ASAT</th>
<th>G6PDH</th>
<th>6GPDH</th>
<th>GPI</th>
<th>MDH</th>
<th>ME</th>
<th>MPI</th>
<th>NH1</th>
<th>NH2</th>
<th>PGM</th>
<th>PK</th>
</tr>
</thead>
<tbody>
<tr>
<td>KMS 1</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>1.2</td>
<td>1.4,5</td>
<td>3</td>
<td>3.4,5</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>KMS 2</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>1.2</td>
<td>1.4,5</td>
<td>3</td>
<td>3.4,5</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>KMS 3</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>1.2</td>
<td>1.4,5</td>
<td>2</td>
<td>3.4,5</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>2</td>
</tr>
<tr>
<td>KMS 4</td>
<td>3</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>1.2</td>
<td>1.6</td>
<td>3</td>
<td>4.5,6</td>
<td>2</td>
<td>1</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>KMS 5</td>
<td>1</td>
<td>3</td>
<td>3</td>
<td>1</td>
<td>4</td>
<td>2.5</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>3</td>
<td>1</td>
<td>3</td>
</tr>
<tr>
<td>KMS 6</td>
<td>2</td>
<td>4</td>
<td>2</td>
<td>1</td>
<td>3</td>
<td>4</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>–</td>
<td>2</td>
<td>3</td>
</tr>
<tr>
<td>KMS 7</td>
<td>4</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>1.2</td>
<td>1.6</td>
<td>3</td>
<td>4.5,6</td>
<td>2</td>
<td>1</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>KMS 8</td>
<td>3</td>
<td>1</td>
<td>2</td>
<td>1.2</td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>4.5,6</td>
<td>2</td>
<td>1</td>
<td>3</td>
<td>1</td>
</tr>
<tr>
<td>KMS 9</td>
<td>2</td>
<td>3</td>
<td>3</td>
<td>1</td>
<td>3</td>
<td>3</td>
<td>2</td>
<td>2</td>
<td>2</td>
<td>–</td>
<td>3</td>
<td>2</td>
</tr>
</tbody>
</table>

ALAT = alanine aminotransferase; ASAT = aspartate aminotransferase; G6PDH = glucose-6-phosphate dehydrogenase; 6GPDH = 6-phosphogluconate dehydrogenase; GPI = glucose-phosphate isomerase; MDH = malate dehydrogenase; ME = malic enzyme; MPI = mannose-phosphate isomerase; NH1 and NH2 = nucleoside hydrolase (inosine); PGM = phosphoglucomutase; PK = pyruvate kinase.

---

**Figure 3.** Relationship among 16 Argentinian *Leishmania* stocks from 12 patients (MAS5a, MAS5b, MRO2a, MRO2b, OLO1, RLS6, AZ3, AAS4, NSS3, HNO3a, HNO3b, FDO4, CFO5, JDM1, and LBC1; a, b mean different isolates from the same patient), a dog (LPO1), and WHO reference strains (M2904, LS94, M4147, M2682, M379, and M2269). The phenogram was built by the UPGMA method from the Jaccard distances calculated.
L. (V.) panamensis should be considered as a unique clade based on the discrete typing unit concept. In this study, only two loci showed different characters between them, reaching a relatively small degree of distance (JD = 0.222). Such a difference may not be enough to separate them as distinct species. Nevertheless, the factors that control the species diversity within the genus Leishmania remain unknown. A better understanding of taxonomic units for Leishmania will come when we understand what controls their distribution.

In a previous study of 208 dogs examined, 41 (19.7%) presented leishmanial lesions, and 14 (45.2%) out of 31 lesions studied intensively were positive for Leishmania amastigotes, but no parasite isolation in vitro was possible. In this work, therefore, we are reporting for the first time the isolation and characterization of Leishmania from dogs. Although the species involved was L. (V.) braziliensis, its zymodeme was not expressed by any of the isolates from humans. To incriminate a reservoir host, the parasites isolated from the reservoirs must be indistinguishable from those of man. This becomes especially important in the case of clades of parasites, such as L. (V.) braziliensis, in which their heterogeneity and plasticity have been demonstrated. Although more isolates from dogs need to be analyzed, this observation adds a point of discussion on whether the animal has a principal role as a reservoir of this disease, even when its prevalence is relatively high in the endemic area. In Minas Gerais State, Brazil, an intraspecific variation among L. (V.) braziliensis from dogs was described. On the other hand, in Três Braços, Brazil, two stocks isolated from dogs were shown to be indistinguishable from 15 isolates from humans by isoenzyme profiles. Furthermore, although L. (V.) braziliensis was found infecting dogs and other domestic animals in the endemic areas of different countries, the stocks characterization had reached, analyzing a limited number of enzymes. Under these circumstances, a generalization about the indistinguishableness among L. (V.) braziliensis isolated from humans and dogs cannot be done as has been suggested. Therefore, in the future, it should be proved in each study area, examining enough materials (human and dog isolates) by suitable techniques for the analysis.

The geographic origin of the zymodesms in the current study suggests that both phytopographical areas, Selva de Transición and Chaco Occidental, are possible transmission places for Leishmania (V.) spp. in Salta Province. Particularly the same zymodeme of L. (V.) braziliensis, KMS 1, was found in both zones. Considering the zoontotic characteristic of ATL, this fact triggers questions such as if the same transmission cycle exists in both areas, is it related to the mobility and the ability of the reservoirs to colonize different environment. Therefore, a better understanding of the disease will be obtained when the transmission cycles have been disclosed in the areas.

Thus, the current data on the zymodesms from humans and dogs will give important information on the causative agents circulating in and around the endemic areas. By performing further analysis using more stocks from humans, it would be possible to disclose the relationships between the clinical features and the zymodeme patterns in the country.

Received June 9, 2004. Accepted for publication August 23, 2004.

Acknowledgments: We are grateful to the physicians and personnel of the Dermatology Ward of Hospital del Milagro and the Dermatology and Otorhinolaryngology Wards of Hospital San Bernardo of Salta City for the recruitment of the patients. We also thank Pedro Rueda, Hilda de Fillipi, Héctor Romero, Marisa Juarez, Federico Ramos, and Ricardo Rossi for their technical support and Patricia Diosque and Marcelo Padilla for their contributions and valuable comments.

Financial support: This study was supported by the Ministry of Education, Science, Culture and Sports of Japan (grant no. 14256002).

Authors’ addresses: Jorge D. Marco, Paola A. Barroso, Manuel Calvopiña, Hídeo Kumazawa, Masataka Korenaga, and Yoshishita Hashiguchi, Department of Parasitology, Kochi Medical School, Kochi University, Kohasu, Oko Cho, Nankoku Shi, Kochi Ken, 783-8505 Japan, Telephone/Fax: +81 88 880 2415. Silvana P. Caijal and Néstor J. Taranto, Instituto de Enfermedades Tropicales, Sede Regional Orán, Universidad Nacional de Salta, Alvarado 751, San Ramón de la Nueva Orán, (4530) Salta, Argentina, Telephone/Fax: +54 3878 421924. Masato Furuya, Institute for Laboratory Animals, Kochi Medical School, Kochi University, Kohasu, Oko Cho, Nankoku Shi, Kochi Ken, 783-8505 Japan, Telephone: +81 88 880 2434, Fax: +81 88 880 2436. María C. Mora and Miguel A. Basombri, Instituto de Patología Experimental, Facultad de Ciencias de la Salud, Universidad Nacional de Salta, Buenos Aires 177, (4400) Salta, Argentina, Telephone/Fax: +54 387 425533. María M. J. Rea, C. Edgardo Borda, Centro Nacional de Parasitología y Enfermedades Tropicales, Facultad de Medicina, Universidad Nacional del Nordeste, Santa Fe 1432, (3400) Corrientes, Argentina, Telephone/Fax: +54 3783 425484.

Reprint requests: Jorge D. Marco, Department of Parasitology, Kochi Medical School, Kochi University, Kohasu, Oko Cho, Nankoku Shi, Kochi Ken, 783-8505 Japan, Telephone/Fax: +81 88 880 2415, E-mail: marcojd@med.kochi-ms.ac.jp.

REFERENCES


